

Pesticidal Plants Used in Masaka District of Uganda

Mwine Julius

The use of synthetic pesticides in developing countries is limited by both the high cost of procuring them and predominance of subsistence holdings. Resultantly, many farmers have to rely on traditional methods of pest control. Among these is the use of pesticidal plant extracts and this paper reports on the findings of a study that undertook to compile an inventory of plants that are used in pest control in one part of the developing world, namely, Masaka District of Uganda. The paper reports that the study found that thirty-six (36) plant species are used. Thirty-five (35) of these were found to belong to twenty-one (21) families. The paper adds that, of these, the Asteraceae family is the most commonly used followed by the Solanaceae family. It was noted that although some of the plants are scientifically well established (e.g. *Azadirachta indica*, *Melia azedarach*, and *Tagetes minuta*), a few are not well known (e.g. *Euphorbia tirucalli*, *Bidens pilosa*, *Vernonia amygdalina*), hence the need for research on them. Finally, it was established that some of the plants are increasingly rare, which highlights need for their conservation.

Key words: Pest Control; Pesticidal Plants; Indigenous Knowledge

Introduction

In many parts of the developing world, the use of synthetic pesticides is limited and, in many cases, entirely nonexistent. This is because their cost is beyond the means of many of the farmers in these areas. Moreover, the predominance of subsistence agricultural holdings in these areas makes the use of such pest control measures uneconomical. Besides, in many instances, the use of these pesticides is maligned, which is aggravated by farmers' ignorance about them. Incidentally, the low utilization of these pesticides is despite the fact that most of these areas lie in tropical, and subtropical, areas, where pests and diseases are profuse throughout the year. Consequently, pests, and the diseases that they transmit, pose one of the major problems affecting agricultural production in these areas. According to FAO (2003), for example, crop loss in these areas exceeds 40%, which is higher than the world average of 30 (Oerke et al., 1994; Oerke and Dehne, 2004).

In view of the constraints hindering the use of modern pesticides, the use of traditional methods of pest control appears to offer a means of overcoming the productivity losses attributable to pests (Rates, 2001; Pei, 2001; Muhammad & Awaisu, 2008). One of these methods is the use of pesticidal plants. Though there is evidence of use of pesticidal plants in pest control as early as the 1500s (Thacker, 2002), the discovery of synthetic pesticides, in the early 1900s, tended to overwhelm their use, because of the advantages associated with the latter. For example, DDT is reported to have had a knockdown effect



on most insects, high persistence in the environment, ease of application and having a broad spectrum (DeLong, 1948; Walker, 2000), advantages that pesticidal plants do not seem to offer.

To be put to extensive use, therefore, pesticidal plants have to be 'rediscovered', documented and evaluated (Muhammad & Awaisu, 2008). Unfortunately, information about them, and their utility, is usually exclusive to a few people, who usually withhold it from others (Ankli et al., 1999). Moreover, even in instances where those that are knowledgeable about such plants pass on their knowledge to others, this dissemination is usually oral, coming with it the limitations of oral communication including limited circulation and the possibility of loss of knowledge due to memory loss and death. Taking the case of Masaka District of Uganda, therefore, this study undertook to compile an inventory of pesticidal plants, to preserve and disseminate the knowledge about them that is available; stimulate further research on them; and promote their conservation.

Methods

The study was carried out in Masaka District, located in central Uganda (between 31° 12' and 32° 06' E; and 0° 48' and 1° 20' S). Data were collected from five sub-counties of the District, namely, Bigasa, Bukoto, Kitanda, Kingo and Kibinge. Masaka District shares the shore of Lake Victoria and is one of Uganda's main agricultural areas. Traversed by the equator, the District receives bimodal rainfall, with an average of 1200mm per annum; and has mild equatorial temperatures, ranging between 22 and 26°C (Britannica online encyclopaedia). Due to the bimodal type of rainfall, the district has two growing seasons, i.e. March to May and October to December, which enables the growing of crops throughout the year. The main crops grown include Bananas (*Musa sp*), Cassava (*Mannibot esculenta*), Maize (*Zea mays*), Coffee (*Coffea sp*) and a range of tropical vegetables, fruits and cereals. Data were collected using questionnaires, interviews and observation. The survey team was made up of an agricultural officer, an agricultural extension worker and field officers. The field officers, who doubled as translators wherever need arose, were selected from the respective sub-counties. In each sub-county, twenty five (25) farmers and one agricultural extension worker were interviewed, making a total of 130 respondents. Each respondent was asked to provide information on any pesticidal plants that they knew. Thus, they provided information on the names of the plants; the pests that they are used to fight; the parts of the plants used; and the method of using the plants against pests. Thereafter, a voucher specimen was collected from the plants identified and deposited at Makerere University Herbarium, for scientific identification.

Findings

The pesticidal plants identified, as well as their families and parts used in controlling pests, are shown in Table 1.

Table 1: Pesticidal Plants Identified

Species name	Local name	Family	Part used
<i>Abrus precatorius</i>	Lusiiti	Papilionaceae	L, S
<i>Allium sativum</i>	Katungulucumu	Alliaceae	Corm
<i>Annona muricata</i>	Kisitaferi	Annonaceae	L, B
<i>Artemisia annua</i>	Artemisia	Asteraceae	L, F
<i>Asparagus africanus</i>	Kadaali	Asparagaceae	Spines
<i>Azadirachta indica</i>	Neem	Meliaceae	L, B R, F
<i>Bidens pilosa</i>	Ssere	Compositae	L
<i>Capsicum frutescens</i>	Kamulari	Solanaceae	F
<i>Carica papaya</i>	Mupaapali	Caricaceae	R, B
<i>Chrysanthemum coccineum</i>	Pyrethrum	Asteraceae	L, F
<i>Citrus aurantifolia</i>	Nimawa	Rutaceae	F, L
<i>Cupressus lusitanica</i>	Kapriposi	Cupressaceae	L, B
<i>Cymbopogon nardus</i>	Mutete	Poaceae	L
<i>Datura stramonium</i>	*	Solanaceae	L, F
<i>Eucalyptus globulus</i>	Kalitunsi	Myrtaceae	L
<i>Eucalyptus grandis</i>	Kalitunsi	Myrtaceae	L
<i>Euphorbia candelabrum</i>	Nkukuulu	Euphorbiaceae	Latex
<i>Euphorbia tirucalli</i>	Nkoni	Euphorbiaceae	Latex, B, ash
<i>Jatropha curcas</i>	Kiryowa	Euphorbiaceae	Sap, F, S
<i>Melia azedarach</i>	Lira	Meliaceae	L, R, B
<i>Mucuna pruriens</i>	Mucuna	Fabaceae	L
<i>Nicotiana tabacum</i>	Taaba	Solanaceae	L
<i>Phoenix reclinata</i>	Mukindo	Palmae	Sap
<i>Phytolacca dodecandra</i>	Luwoko	Phytolaceae	L, F
<i>Ricinus communis</i>	Nsogasoga	Euphorbiaceae	S
<i>Schinus molle</i>	*	Anacardiaceae	L, F
<i>Solanum lycopersicum</i>	Enyaanya	Solanaceae	F
<i>Tagetes minuta</i>	Kawunyira	Asteraceae	L
<i>Tithonia rotundifolia</i>	Ekimyula	Asteraceae	F, L
<i>Cannabis sativa</i>	Njagga	Cannabaceae	L, S, F
<i>Lantana camara</i>	Kayukiyuki	Verbenaceae	L
<i>Tephrosia vogelii</i>	Muluku	Fabaceae	L
<i>Cupressus sempervirens</i>	Ssederu	Cupressaceae	S, L
*	Olukomba	*	L, F
<i>Vernonia amygdalina</i>	Omululuza	Compositae	L
*	Mutanjoka	*	L

Legend: B=Bark; F=Fruit; L=Leaves; R=Roots; S=Seeds; *Not identified

Table 1 indicates that thirty-six (36) pesticidal plants were identified. Of these, plants from the Asteraceae and Solanaceae families were commonest (four from each of the families), followed by those from the Euphorbiaceae family (three). Two of the families identified in the Table belonged to the grasses subdivision (i.e. Palmaceae and Poaceae) while the rest belonged to dicotyledonous subdivisions. Leaves were reported to be the most commonly used part of the plants, followed by fruits, seeds, barks, roots and sap. Nonetheless, several intersections of parts used were reported and *Azadirachta indica*, *Melia azedarach*, *Cannabis sativa* and *Jatropha curcas* were reported to be particularly potent. The pests controlled by the plants identified and methods of formulating the plants are shown in Table 2.

Table 2: Pests Controlled by the Plants Identified and Methods of Formulating the Plants

Species name	Pest/ Disease treated	Mode of formulation
<i>Abrus precatorius</i>	Worms	Water extract
<i>Allium sativum</i>	Field and storage pests	Trap crop and water extract
<i>Annona muricata</i>	Fungicidal properties and insects	Water extract
<i>Artemisia annua</i>	Mosquitoes and flies	Water extract
<i>Asparagus africanus</i>	Birds, moths and bats	Physical trap/thorns
<i>Azadirachta indica</i>	Most insects	Water extract/crashed seeds
<i>Bidens pilosa</i>	Aphids	Water extract
<i>Capsicum frutescens</i>	Cut worms, ants, snails and storage pests	Water extract/ crashed seeds
<i>Carica papaya</i>	Blight and animal worms	Leaf water extract and seeds
<i>Chrysanthemum sp</i>	Most pests	Oil extract form leaves
<i>Citrus aurantifolia</i>	Insect pests	Water extract and trap crop
<i>Cupressus lusitanica, sempervirens</i>	Storage pests	Physical mixture and leaf oil
<i>Cymbopogon nardus</i>	Lepidoptera pests, beetles and aphids	Trap crop, repellent and oil extract
<i>Datura stramonium</i>	Insects pests	Water extract
<i>Eucalyptus globulus</i>	Storage pests	Water extract, physical mixture and oil extract
<i>Eucalyptus grandis</i>	Storage pests	Water extract, physical mixture and oil extract
<i>Euphorbia candelabrum</i>	Termites and cutworms	Latex spray and root effect
<i>Euphorbia tirucalli</i>	Aphids, safari ants and cutworms	Latex spray and ash dusting/ physical mixture
<i>Jatropha carcas</i>	Insect pests and animal worms	Latex spray, water extract and crushed seeds/ oil
<i>Melia azedarach</i>	Worms and most insects	Water extract
<i>Mucuna pruriens</i>	Most insects	Water extract
<i>Nicotiana tabacum</i>	Storage pests, soil pests, domestic pests and snake repellent	Water extract and smoke (fumigant)
<i>Phoenix reclinata</i>	Storage pests	Ash
<i>Phytolacca dodecandra</i>	Snails, insect pests and fungi	Water extract

Table 2 Continued

Species name	Pest/ Disease treated	Mode of formulation
<i>Ricinus communis</i>	Storage pests, domestic pests and animal worms	Water extract and crushed seed /oil
<i>Schinus molle</i>	Worms and storage pests	Water extract and crushed seeds
<i>Solanum lycopersicum</i>	Aphids, thrips and weevils	Water extract and repellent
<i>Tagetes minuta</i>	Nematodes and most insects	Water extract, repellent and trap
<i>Tithonia rotundifolia</i>	Nematodes and Aphids	Water extract
<i>Cannabis sativa</i>	Storage pests, insect pests, Coccidiosis and bactericide	Water extract, smoke and trap
<i>Lantana camara</i>	Insect pests, Storage pests	Water extract and dry leaf mixture
<i>Tephrozia vogelii</i>	Insect pests, Ticks, Fungi, mites and moles	Water extract and roots
<i>Cupressus sempervirens</i>	Bean weevils and house flies	Water extract and repellent
<i>Orukomba</i>	Insects and Fungi	Water extract and repellent
<i>Vernonia amygdalina</i>	Insect pests, Malaria parasites and worms	Water extract
<i>Mutarjoka</i>	Worms	Water extract

The results in Table 2 indicate that the plants that were identified were used against a wide range of pests, including weevils, storage pests, caterpillars, insects and field pests. They also indicate that water extraction was reported to be the most commonly used mode of formulating the plants. Others included the use of the plants as pest repellents; crushing of their seeds and extraction of oil; and using them as pest snares. It may be noted that many of the plants could be formulated in more than one way.

Discussion

The study identified thirty-six (36) plant species that are used in pest control in Masaka District (see Table 1). Thus, it corroborates the findings of earlier researchers (e.g. Rates, 2001; Pei, 2001; Muhammad and Awaisu, 2008) indicating that the use of traditional methods of pest control appears to offer a means of overcoming the productivity losses attributable to pests in various parts of the tropics and subtropics. Whereas some of these species belonged to well-known families (e.g. *Azadirachta indica*, *Melia azedarach*, *Jatropha curcas*, *Tagetes minuta*, *Tithonia rotundifolia*, *Chrysanthemum sp* (Isman, 2006)), the efficacy of some of the species (e.g. *Euphorbia tirucalli*, *Euphorbia candelabrum*, *Bidens pilosa*, *Vernonia amygdalina*) is not well established in the literature. Incidentally, although the farmers interviewed could identify the pests controlled by the plants identified, they could not pinpoint them at the family level, which caused the lumping of the pests identified in amorphous categories that may not help in the drawing of comparisons and scientific conclusions. This does not only reaffirm the view that pesticidal plants have to be ‘rediscovered’, documented and evaluated if they are to be put to extensive use (Muhammad and Awaisu, 2008) but also that there is need for further research into the pesticidal nature of the plants identified, notwithstanding the current study. Specifically, there is need for efficacy studies, to recommend the plants for extensive use in pest control.

Leaves were found to be the most commonly used part of the pesticidal plants identified (see Table 2). This is in agreement with earlier researchers (e.g. Ssegawa & Kasenene, 2007; Kamatenesi-Mugisha et al., 2007; Maregesi et al., 2007), who found leaves to be the part of plants that is most commonly used for medicinal purposes. A possible explanation for this is that, for defensive purposes, plants tend to develop and deposit secondary substances—like alkaloids (Dethier, 1980), Tanins (Bernays, 1981), Phenols (Palo, 1984) and Monoterpins (Schutte, 1984), some of which are toxic to some pests—in their body parts, especially leaves because they are more exposed to pests (Gatehouse, 2002). In fact, these are the substances used in the making of pesticides of a botanical nature. This suggests that researchers delving into pesticidal plants should pay special attention to leaves.

The interviews conducted revealed that some of the pesticidal plants (e.g. *Abrus precatorius*, *Phoenix reclinata*, and *Euphorbia candulubrum*) are increasingly scarce while others can no longer be obtained from the area. Thus, the study gives credence to the fears raised by Hedberg (1993) and Cox (2000), that these plants are tending towards extinction unless information about their utility is documented and the plants conserved.

References

- Ankli, A., Sticher, O. & Heinrich, M., 1999. Medical ethnobotany of the Yucatec Maya: Healers’ consensus as a quantitative criterion. *Economic Botany*, 53, pp. 144-160.
- Bernays, E. A., 1981. Plant tannins and insect herbivores: an appraisal. *Ecological Entomology*, 6, pp. 353-360.
- Britannica Online Encyclopaedia, 2008. *Uganda climate*. [Online] Available at: <http://www.britannica.com/eb/article-37600>. [Accessed 2008].
- Cox, P. A., 2000. Will tribal knowledge survive the millennium? *Science*, 287, pp. 44-45.
- Delong, D. M., 1948. DDT and the insect problem. *The Quarterly Review of Biology*, 23 (1) pp. 59-160.

- Dethier, V. G., 1980. Evolution of receptor sensitivity to secondary plant substances with special reference to deterrents. *American Naturalist*, (115) 45.
- FAO, 2003. Crop protection in the context of Agricultural development. In: *Report of the First External Review of the System-wide Programme on Integrated Pest Management (SP-IPM)*
- Gatehouse, J. A., 2002. Plant resistance towards insect herbivores: a dynamic interaction. *New Phytologist*, 156, 145-169.
- Hedberg, I., 1993. Botanical methods in Ethnopharmacology and the need for conservation of medicinal plants. *Journal of Ethnopharmacology*, 38, 121-128.
- Isman, M. B., 2006. Botanical insecticides, deterrents and repellents in modern agriculture and an increasingly regulated world. *Annu Rev Entomol*, 51, 45-66.
- Kamatenesi-Mugisha, M., Oryem-Origa, H. & Olwa-Odyek, 2007. Medicinal plants used in some gynaecological morbidity ailments in Western Uganda. *African Journal of Ecology*, 45, 34-40.
- Maregesi, S. M., Ngassapa, O. D., Pieters, L. & Vlietinck, A. J. 2007. Ethnopharmacological survey of the Bunda District, Tanzania: plants used to treat infectious diseases. *Journal of Ethnopharmacology*, 113, 457-470.
- Muhammad, B. Y. & Awaisu, A., 2008. The need for enhancement of research, development and commercialization of natural medicinal products in Nigeria: lessons from the Malaysian experience. *African Journal of Traditional Complementary and Alternative Medicines*, 5, 120-130.
- Oerke, E. C., Dehne, H. K., Schonbeck, F. & Weber, A., 1994. Crop production and crop protection: estimated losses in major food and cash crops. *Malaria Foundation International*, 808.
- Oerke, E., C. & Dehne, H., W., 2004. Safeguarding production losses in major crops and the role of crop protection. *Crop Protection*, 23, 275-285.
- Palo, R. T., 1984. Distribution of birch (*Betula* SPP), willow (*Salix* SPP), and poplar (*Populus* SPP) secondary metabolites and their potential role as chemical defence against herbivores. *Journal of Chemical Ecology*, 10, 499-520.
- Pei, S. J., 2001. Ethnobotanical approaches of traditional medicine studies: Some experiences from Asia. *Pharmaceutical Biology*, 39, 74-79.
- Rates, S. M. K., 2001. Plants as source of drugs. *Toxicon*, 39, 603-613.
- Schutte, H. R., 1984. Secondary plant substances: Monoterpenes. *Progress in Botany*, 119-139.
- Ssegawa, P. & Kasenene, J. M., 2007. Medicinal plant diversity and uses in the Sango bay area, Southern Uganda. *Journal of Ethnopharmacology*, 113, 521-540.
- Thacker, J. R. M., 2002. *An introduction to arthropod pest control*. Cambridge: Cambridge University Press.
- Walker, K., 2000. Cost-comparison of DDT and alternative insecticides for malaria control. *Medical and Veterinary Entomology*, 14, 345-354.

